

Nanostructured Sensors for Detection of Heavy Metals: A Review

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ABSTRACT: Heavy metal pollution is one of the most serious environmental problems, which undermines global sustainability. Many efforts have been made to develop portable sensors for monitoring heavy metals in the environment. Incorporation of nanomaterials and nanostructures into sensors leads to significant improvement in the performance of devices in terms of sensitivity, selectivity, multiplexed detection capability and portability. In addition, small molecules, DNA, proteins and bacteria have been integrated with inorganic materials to selectively bind heavy metals as the molecular recognition probes. This review presents a recent advance in optical, electrochemical and field-effect transistor sensors for heavy metal detection. The optical sensors are focused on colorimetric, fluorescent, surface-enhanced Raman scattering and surface plasmon resonance devices. In addition, optofluidic devices which integrate optical components with microfluidic chips are discussed. Furthermore, nanoparticle-modified electrodes, microelectrode (or nanoelectrode) arrays and microfluidic electrochemical sensors are



summarized. This paper highlights the strategies for design of nanostructured sensors and the benefits from the use of nanomaterials and nanostructures.

KEYWORDS: Heavy metal, Optical sensor, Electrochemical sensor, Nanomaterial, Biosensor, Microfluidics, Optofluidics

Heavy metals enter the environment due to increasing industrial activities. Heavy metals such as Hg, As, Pb, and Cd are highly toxic and carcinogenic even at a trace level.^{1,2} They are nonbiodegradable and can accumulate in the food chain, which poses a severe threat to the environment and human health. Heavy metal pollution becomes a concern for global sustainability. It is therefore essential to monitor heavy metals in the environment, drinking water, food, and biological fluids. Conventional methods for heavy metal measurement include atomic absorption spectroscopy, inductively coupled plasma/ mass spectrometry, inductively coupled plasma/atomic emission spectrometry, ultraviolet-visible spectroscopy, etc.³⁻⁶ Although these techniques are highly sensitive and selective, they require tedious sample preparation and preconcentration procedures, expensive instruments, and professional personnel.⁷ Moreover, they cannot be used as portable devices for on-site detection. In contrast, sensors have great potential in high throughput detection of multiple heavy metals on-site. Rapid development of nanotechnology has provided new opportunities for improving the performance of sensors in terms of sensitivity, limit of detection, selectivity, and reproducibility, and also enabled miniaturization with assistance of lab-on-chip (LOC) technology.^{8,9}

This paper reviews the recent progress in nanostructured sensors for heavy metal detection. Sensors are summarized based on different signal transduction mechanisms, including optical, electrochemical, and field-effect transistor (FET) sensors. The optical sensors include fluorescent, colorimetric, surface plasmon resonance (SPR), and surface-enhanced Raman scattering (SERS) sensors. Special emphasis is put on the role of nanomaterials and nanoarchitecture in sensors. In addition, this paper highlights the integration of sensors with microfluidic chips, which is an important direction toward applicability of sensing devices to real-world samples.

■ FLUORESCENT SENSORS

Fluorescence sensing is based on analyte-induced changes in the physicochemical properties of fluorophores including fluorescence intensity, lifetime, and anisotropy, which are related to charge transfer or energy transfer processes.¹⁰ Researchers have put a spotlight on the Förster resonance energy transfer (FRET) process when developing fluorescent sensors for heavy metal detection. FRET occurs through the dipole–dipole interaction between an energy donor and an acceptor, following a $1/d^6$ separation distance dependence.^{11–13} The effective interaction distance (Förster distance) typically is ~6 nm. Typically, FRET requires the spectral overlap between the emission band of donor and the absorption band of acceptor.¹⁴ Ono has pioneered the development of FRET sensors for heavy metal detection.^{15,16} Ono has linked an organic dye (fluorophore) and a quencher to two ends of a molecular beacon to form a FRET sensor, as shown in Figure 1. The presence of Hg^{2+} ions induced a hairpin structure due to the formation of the $T-Hg^{2+}-T$ sandwich structure, which brought the quencher close to the fluorophore and thus enabled the FRET process, leading to fluorescence quenching

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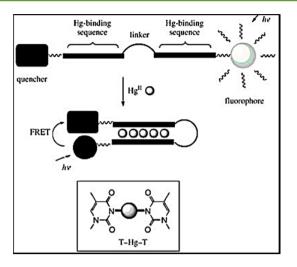


Figure 1. Sketch showing the molecular beacon-based FRET sensor; the presence of Hg^{2+} ions leads to quenching of fluorescence emission of fluorescein.¹⁶.

of the organic dye. This sensor achieved a limit of detection (LOD) of 40 nM toward Hg^{2+} in a buffer solution and excellent selectivity toward Hg^{2+} ions coexisting with other metal ions. Ono's work has opened a new avenue for development of fluorescent heavy metal sensors.

Organic dyes usually are used as the fluorophores in FRET sensors due to the commercial availability of toolboxes, wellestablished labeling protocols, and small size that minimizes possible steric hindrance. However, organic dyes suffer from photobleaching and have narrow excitation spectra.^{17,18} Inorganic quantum dots (QDs), graphene oxide (GO), carbon dots, metallic clusters, and upconversion luminescent nanoparticles (NPs) have been emerging as alternative fluorophores due to their unique properties. For example, QDs possess broad excitation profiles, narrow and symmetric emission spectra, high photostability, high quantum efficiency, size-dependent fluorescence emission peaks, and potential multiplexed detection capability.¹⁹⁻²³ Carbon dots and graphene QDs are free of heavy metal elements, possess excellent biocompatibility, and have attractive surface functionality.^{24–26} Owing to their good biocompatibility, polymer nanomaterials and carbon dots are of interest to fluorescent sensors used in in vivo and in vitro detection of heavy metals. In addition, upconversion NPs are explored for in vivo sensing due to their low background interference by the near-infrared excitation source.²⁷

When the organic quencher is replaced with a gold NP in a fluorophore-quencher system, the energy transfer mechanism is dependent on the plasmonic nature of the Au NP. If the Au NP exhibits surface plasmon resonance (SPR), the energy transfer from the fluorophore (an organic dye or a quantum dot) to the gold NP follows the FRET mechanism.³⁰ When the Au NP has no SPR, the energy transfer mechanism is switched from FRET to nanometal surface energy transfer (NSET).³⁰ Unlike FRET, NSET is originated from the dipole-induced interband electronic transition in a thin layer of a semi-infinite metal hemisphere, which does not require the resonant interaction between electrons.^{30,31} The NSET process follows a $1/d^4$ separation distance dependence and exhibits a much longer effective interaction distance (~30 nm).^{31–34} Au NPs are excellent fluorescence quenchers over a broad range of wavelengths and long distances in comparison with organic acceptors.^{30,32,33} Au NP quenchers have several appealing

features.^{30,35–37} (i) Au NPs have no defined dipole moment as do organic molecules. Hence, the energy transfers in any orientation relative to the Au NPs. (ii) Au NPs have large absorption cross sections near the plasmon resonance. (iii) Au NPs have no fluorescence emission, avoiding the crossexcitation that usually occurs in organic dyes. (iv) Au NPs have low toxicity.

Recently a QD–DNA–Au NP ensemble was created by utilizing the NSET mechanism.³⁸ As shown in Figure 2, the

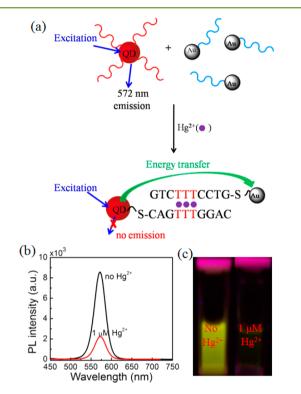


Figure 2. (a) Sketch showing the QD–DNA–Au NP ensemble sensor for Hg^{2+} detection. (b) Fluorescence emission spectra. (c) Photograph of the QD/DNA/Au NP solution under 365 nm laser excitation in the absence and presence of 1 μ M Hg^{2+,38}

complementary oligonucleotide strands were linked to the QDs and Au NPs, respectively. When Hg²⁺ ions were present in the aqueous solution that contained the oligonucleotide-conjugated QDs and Au NPs, Hg²⁺ ions selectively bound to thymine in the oligonucleotides, forming the DNA helix. As a result, the QDs and Au NPs were brought into a close proximity, leading to NSET from the QD to Au NPs. Consequently, the fluorescence emission of the QD was quenched by the Au NPs. This nanosensor exhibited a LOD of 0.4 and 1.2 ppb toward Hg²⁺ in the buffer solution and in river water, respectively, and excellent selectivity toward Hg²⁺ over various metal ions. In addition, a Rhodamine B-Au NP-based NSET probe has been reported for monitoring mercury in water, fish, and contaminated soil. This inexpensive and battery-operated sensor exhibited excellent sensitivity (2 ppt) and selectivity for Hg²⁺ over Cu²⁺ and Pb²⁺.³⁹

Graphene oxide (GO) is an interesting fluorescence quencher alternative to the Au NPs in NSET sensors.^{40–42} GO is inexpensive and can be produced massively. It has excellent water solubility and a facile surface-functionalization feature due to its inherent oxygen-containing functional groups.^{43–46} Moreover, it has super fluorescence quenching capability. Hence, GO is attractive for sensors based on energy transfer.^{47–51} For example, a fluorescent QD–aptamer–GO sensor was designed based on the NSET mechanism in order to detect Pb^{2+} ions (Figure 3).⁴⁰ The QD-labeled aptamer, which

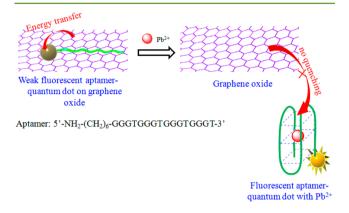


Figure 3. Sketch showing a "turn-on" fluorescent QD-aptamergraphene oxide sensor for Pb^{2+} detection.⁴⁰

was employed as the molecular recognition probe, was physically adsorbed onto the GO surface via the strong $\pi - \pi$ stacking interaction between the single-stranded DNA and GO. Fluorescence of the QDs adsorbed on the GO was quenched due to the NSET from the QDs to GO. The presence of Pb²⁺ in the assay led to the formation of a G-quadruplex/Pb²⁺ complex that had very weak affinity with the GO sheet. Consequently, the QDs conjugated with the G-quadruplex/ Pb²⁺ complex was detached from the GO sheets, which turned on the fluorescence emission of QDs. This sensor exhibited a LOD of 90 pM and excellent selectivity over a wide range of metal ions. In addition, a GO–DNAzyme-based biosensor was developed for detection of Pb^{2+,52} in which the GO also acted as a quencher for an organic fluorophore.

Besides conventional fluorescent sensors based on FRET and NSET, chemiluminescent sensors have attracted interest for detection of heavy metals.^{53,54} In the chemiluminescent resonant energy transfer (CRET) sensors, luminescence of the donor is excited via a chemical reaction without the need of an external light source, which can simplify the design of the sensor system and reduce the noise background.

In addition to the energy transfer processes, the electron transfer process has also been employed for designing fluorescent sensors for heavy metal detection.^{47,55,56} The electron transfer proceeds via different mechanisms including the Dexter interaction, intramolecular photoinduced electron transfer, and interfacial electron transfer. The rate constant of electron transfer exponentially decays with the distance from the nuclei, and the electron transfer is efficient at separation distances less than 1 nm. $^{57-59}$ By utilizing the electron transfer mechanism, a QD-based fluorescent sensor was developed for simultaneous detection of Hg²⁺ and Ag^{+,55} This sensing assay initially contained the free-standing nucleic acid-functionalized QDs, which emitted fluorescence under light excitation. When Hg^{2+} or Ag^+ appeared in the assay, the metal ions were sandwiched in the rigid hairpin structure of DNA, which enabled electron transfer from the QD to Hg²⁺ ions along the duplex DNA channel, quenching the fluorescence of the QD.

It is worth noting that GO can act as not only a quencher (acceptor)⁴⁰ but also a fluorophore (donor).^{43,56} Unlike QDs such as CdS, CdSe, and PbS, GO not only contains a nontoxic

element but also possesses inherent carboxylic acid that can be easily conjugated to amine-functionalized DNA and antibody molecules. Therefore, GO is a promising alternative to QDs as a fluorophore in sensors. For example, the aptamer-functionalized GO has been employed as a fluorophore in a sensor based on the electron transfer mechanism for label-free detection of Hg^{2+} in the aqueous solution.⁵⁶ Although this GO-based sensor was designed and constructed in a very simple way, it was a very effective sensor. The sensor showed a LOD as low as 0.92 nM for Hg^{2+} in an aqueous solution.

PLASMONIC SENSORS

Nanoparticle-Based Colorimetric Sensors. SPR arises from collective oscillation of free conduction electrons in the noble metal in resonance with the incident electromagnetic radiation.^{60,61} For example, 20 nm sized monodispersed Au NPs in the aqueous solution exhibit a strong SPR peak in the absorption spectrum, and the aqueous solution shows a "red wine" color. When the Au NPs aggregate to some degree, the color of the solution will change due to a shift of the SPR peak. On the basis of this principle, colorimetric sensors are constructed for detection of various analytes.⁶²⁻⁶⁷ This type of sensor provides direct, visual, and rapid detection of analytes, which minimizes the costs. Hupp has pioneered the development of colorimetric sensors for heavy metal detection.⁶ ' So far, the colorimetric detection method has been used for monitoring various heavy metals such as Hg^{2+} , Pb^{2+} , Cu^{2+} , and As^{3+} .^{69–78} Liu et al. have demonstrated the colorimetric detection of Hg^{2+} using the oligonucleotide functionalized Au NPs (Figure 4).⁷⁶ The selectivity toward Hg^{2+} was attributed to

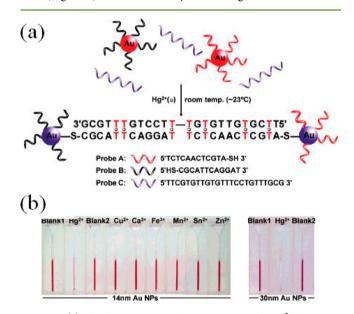


Figure 4. (a) Sketch showing a colorimetric sensor for Hg^{2+} detection. (b) Color response of the 14 nm (bottom left) and 30 nm (bottom right) Au NPs after addition of metal ions (Hg^{2+} , Cu^{2+} , Ca^{2+} , Fe^{3+} , Mn^{2+} , Sn^{2+} , Zn^{2+} ; $10 \ \mu M$ each).⁷⁶

selective binding of Hg²⁺ to the T–T mismatches, leading to aggregation of Au NPs. Aggregation of Au NPs resulted in a color change due to coupling of SPR of the Au NPs in close proximity. This sensor reached a LOD of 3 μ M toward Hg²⁺. Although the colorimetric detection method is simple and rapid, its LOD is relatively high and requires the preconcentration step to detect trace metals. In addition, it remains a

challenge to apply the colorimetric sensors to real-world sample matrices such as human blood and plasma.

Chip-Based SPR Sensors. The position and intensity of the SPR peak are sensitive to change in the surrounding environment (i.e., refractive index) and coupling with the nearby plasmonic field.^{79–81} According to this principle, transmission localized surface plasmon resonance (T-LSPR) spectroscopy can be performed from a plasmonic substrate that can selectively capture metal ions. The resulting SPR peak varies upon the change in the refractive index during capture of heavy metals on the substrate.⁸⁰

SERS SENSORS

SERS sensors have been extensively used for chemical and biological sensing and medical diagnostics,^{82,83} but only a few papers have reported the heavy metal detection.⁸⁴⁻⁹¹ Although SERS as a molecular spectroscopy can provide the spectral fingerprint signatures of analytes, it is unable to directly detect heavy metal ions. A solution to this problem is to functionalize plasmonic nanostructures with an organic ligand that binds specifically to heavy metal ions. As mentioned above, aggregation of Au NPs can be used for colorimetric detection. On the other hand, aggregation of Au NPs can lead to coupling of plasmonic fields of neighboring NPs, forming "hot spots" for SERS enhancement. On the basis of this principle, Chen et al. have developed a SERS sensor for detection of As³⁺. In this sensor, glutathione, which can selectively bind to As³⁺ ions through the As-O bond, was conjugated onto the Ag NPs, and 4-mercaptopyridine (4-MPY) was used as the Raman reporter.⁸⁵ In the presence of As^{3+} ions, binding of As^{3+} with glutathione induced the aggregation of Ag NPs, which made the Raman reporter molecules located in "hot spots", enhancing the SERS signal from 4-MPY. This SERS sensor was simple but achieved a LOD as low as 0.76 ppb with excellent selectivity over various metal ions. However, there is still concern with the stability and reproducibility of the colloid-based SERS sensors.

SERS provides both the information regarding the presence of the chemical element and its chemical form, which is a critical aspect in the toxicological study of metal ions because different complex forms of heavy metals exhibit distinct toxicity toward human or animals. Accordingly, SERS has been employed for direct detection of UO^{2+} , TcO^{4-} , CrO_4^{2-} , NpO_2^+ , NpO_2^{2+} , WO_4^{2-} , etc.⁸⁸⁻⁹⁰ Irudayaraj et al. have demonstrated SERS sensing and imaging of toxic chromate in bacteria.⁹¹ Their results showed that SERS exhibited high sensitivity and was able to distinguish two stable valence forms of chromate in cells.

OPTOFLUIDIC SENSORS

Optofluidic devices refer to the miniaturized microfluidic systems that integrate optical sensors with microfluidics, which brings a number of unique characteristics.^{92–96} Optofluidic devices enable real-time detection within a minimized sample volume and facilitate multiplexed detection of analytes in a single chip simultaneously.⁹⁶ In microfluidic SERS devices, the SERS substrates are incorporated into the microfluidic channels in two ways.⁹² (i) The metal NPs, which act as the SERS substrates, are suspended in the fluid passing through the channel. (ii) A metal nanostructure is fixed on the bottom of a microfluidic channel as the SERS substrate. Choo has combined the droplet-based microfluidics with the SERS transducer for detection of Hg²⁺ (Figure 5).⁹⁷ The Rhodamine

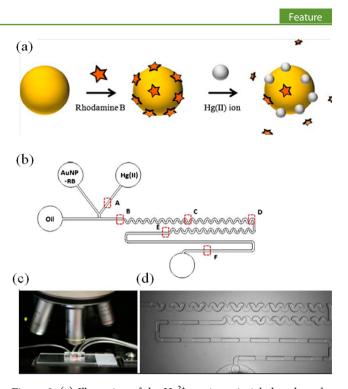


Figure 5. (a) Illustration of the Hg^{2+} sensing principle based on the replacement of the dye molecules with the reduced Hg^{2+} ions on the surface of Au NPs. (b) Microchannels for droplet generation. (c) Microfluidic chip under the microscope. (d) Photograph of the device in operation. ⁹⁷

B molecules were preadsorbed on the Au NP surface, which can be replaced by Hg²⁺ due to stronger affinity between the Au NPs and Hg²⁺ ions. As a result, the SERS signal from Rhodamine B varied as a function of the Hg²⁺ concentration. Another interesting feature of the microfluidic SERS devices is their ability to monitor and separate heavy metals.⁹⁸ However, the number of active sites and accessibility of analytes to the SERS active sites are reduced in the microfluidic SERS devices. This challenge can be addressed by either incorporation of photonic crystals into the device or preconcentration of analytes.^{99,100} Another approach is to use an optofluidic resonator for SERS excitation in the microfluidic environment.^{101,102} The high-intensity field at the optical resonator can function as a high-power excitation source for SERS and, thus, enhance the Raman signal.

ELECTROCHEMICAL SENSORS

Compared to optical sensors, the sensing signal of electrochemical sensors is collected through conducting wires instead of optical detectors. Hence, electrochemical sensors can be easily packed into a compact system. In addition, because heavy metals have the defined redox potential, the selectivity toward specific heavy metal ions can be achieved by bare electrodes without the need of a molecular recognition probe. Several techniques are employed in electrochemical sensing, including voltammetry, amperometry, potentiometry, impedemetry, and conductometry.^{103–105} In particular, the anodic stripping voltammetry (ASV) method is readily amendable for determination of heavy metals. ASV analysis typically involves two steps:^{106–108} (i) electrochemical deposition or accumulation of heavy metals at a constant potential to preconcentrate the analyte onto the electrode surface, and (ii) stripping or dissolution of the deposited analyte from the electrode surface. The mercury-based electrode was the first one that received attention for electrochemical detection of heavy metals because it brought high sensitivity, good reproducibility, and a wide cathodic potential range for heavy metal detection.^{109,110} However, owing to its high toxicity, the mercury-based electrode has been replaced by the environmentally friendly bismuth electrode that exhibits similar electrochemical behavior in many aspects.^{111,112} Unfortunately, the bismuth electrode has a relatively narrow cathodic potential range and suffers from the instability in air due to its natural oxidation.¹¹³ Besides the mercury- and bismuth-based electrodes have also been used in ASV analysis of heavy metals.^{114,115} A detection level of 68 nM has been achieved in 0.1 M KNO₃ using a boron-doped diamond electrode with the differential pulse voltammetric technique.¹¹⁴

Although bulk electrodes have been used in ASV analysis of heavy metals,^{114–118} the LOD and the sensitivity of bulk electrodes cannot meet the need for detection of trace heavy metals. Bulk electrodes (or macro-electrodes) have several problems: (i) a large overpotential required for analyte deposition, (ii) a high stripping potential needed for analyte detection, (iii) the interference of other coexisting metals, and (iv) the interference due to supporting electrolyte ions. An effective way to solve these problems is surface-modification of bulk electrodes with nanoparticles.^{119,120} Another route is to use the micro- or nanoelectrodes.

Nanoparticle-Modified Electrodes. Nanoparticle-modified electrodes possess higher surface area, improved electron transfer rate, increased mass-transport rate, lower solution resistance, and higher signal-to-noise ratio.^{107,119,121} Au NPs have been used to modify bulk electrodes.^{122,123} It has been demonstrated that Au NP-modification of glassy carbon electrodes eliminated the memory effect and interferences of other ions from intermetallic compounds. Moreover, the Au NP-modified glassy carbon electrode significantly lowered the LOD toward Hg^{2+116} It has also been reported that the sensitivity of Au NP-modified electrodes was an order of magnitude higher than that of the macro-electrode counterpart when used for As3+ detection.¹²⁴ Also, the stripping peak of As³⁺ was narrower and more symmetric for the Au NPmodified electrode, which mitigated the effect of copper interference on the arsenic detection. Recently, a Bi nanoparticle-modified electrode was employed for heavy metal detection.¹²⁵ The results showed that the sensitivity and LOD of the electrode were improved with a decrease in particle size of the Bi nanopowder, which was ascribed to a higher electroactive surface area.

In addition, carbon nanomaterials such as carbon nanotubes (CNTs), carbon nanofibers, and graphene have been explored as the electrode materials for detection of heavy metals.^{126–130} For electrochemical sensing applications, CNTs and graphene have many advantages including a large surface area, small size, excellent electron transfer ability, and easy surface-modification. In addition, CNTs have been recognized as excellent sorbents for heavy metal ions.¹³¹ Therefore, it is promising to construct electrochemical sensors using the CNT- or graphene-modified electrodes for heavy metal detection because they are capable of simultaneous preconcentration and detection of multiple heavy metal ions. A single-walled CNT electrode was used to simultaneously measure the Cd²⁺ and Pb²⁺ ions in a 0.02 M HCl solution with the square-wave stripping voltammetry method. It showed a LOD of 2.2 ppb for Cd²⁺ and 0.6 ppb for

Pb²⁺. However, the stripping peak current dropped severely after day-to-day operation.

Because the bare CNTs are hydrophobic, the CNTs are typically modified with functional groups to make them hydrophilic. The surface-functionalization also alters the affinity with heavy metals, which could be utilized for selective detection of heavy meals. For example, the CNTs covalently functionalized with thiacalizarene (TCA) were immobilized on a glass carbon electrode to detect trace Pb^{2+} ions with the differential pulse ASV method.¹³² In this electrode, the CNTs enhanced the electron transfer, and the TCA enabled the selective accumulation of Pb²⁺ ions at the electrode due to its excellent selective recognition. As a result, this electrode exhibited excellent selectivity and high sensitivity for electrochemical detection of Pb^{2+} ions over a range from 2×10^{-10} to 1×10^{-8} M with a LOD of 4×10^{-11} M. In addition, the cysteine-modified CNTs that were cast on the glassy carbon electrode were used for ASV measurement of Pb²⁺ and Cu²⁺.¹³³ The sensor utilized the high affinity of cysteine toward some heavy metals and exhibited the LOD of 1 ppb for Pb²⁺ and 15 ppb for Cu²⁺.

Nanostructured electrodes have also been constructed by the composites that combine two to three materials together to provide synergistic contribution from individual components.^{134–137} Nanostructured metal oxides are typically not involved in the redox process in heavy metal detection, but they can combine with other active materials to improve the sensing performance.¹³⁴ Polymers have also been integrated with CNTs and graphene for heavy metal detection.¹³⁰

Microelectrode and Nanoelectrode Arrays. Even tracelevel (ppb) heavy metals pose a threat on humans and environment. This places a demand on low LOD for sensors. An effective way to improve the LOD and sensitivity of electrochemical sensors is to employ microelectrode arrays (MEAs) and nanoelectrode arrays (NEAs).^{138–140} MEA and NEA refer to a collection of microelectrodes and nanoelectrodes. They have advantages over conventional macro-electrodes, 138,141,142 including (i) a high signal-to-noise ratio because the noise level depends on the active area of the electrode, while the signal depends on the total geometric area of the diffusion field, (ii) no need for convection due to the enhanced mass transport, (iii) a small current that enables stripping analysis to be performed in a high resistive media, which makes the supporting electrolyte unnecessary and hence reduces the interference effects, and (iv) small overpotential required for analyte deposition and stripping.

An array of 256 gold microelectrodes in a diameter of 5 μ m with an interelectrode distance of 100 μ m was used for detection of Hg^{2+.143} This microelectrode array exhibited excellent anti-interference capability due to the short deposition time and low overpotential required for ASV analysis. ASV measurement of Hg²⁺ ions usually encounters the problem of interference from coexisting chemical species such as chloride due to the formation of insoluble calomel (Hg₂Cl₂). It was interesting that this microelectrode array was able to measure Hg^{2+} in 0.1 and 1 M chloride media in the range from 5×10^{-8} to 1×10^{-6} M, avoiding the interference of the calomel formation on the voltammetric peak of interest. In addition, Feeney et al. have developed an Ir-based microelectrode array consisting of 25 microdiscs in a diameter of 10 μ m and an interelectrode distance of 100 μ m, which was used for the determination of 20–100 μ g/L Cd in the aqueous solution. Furthermore, the vertically aligned CNT array was also used for

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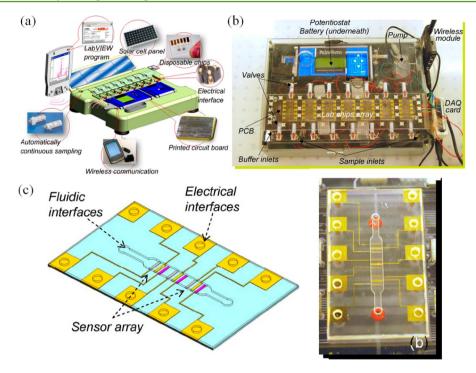


Figure 6. (a) Sketch showing a polymer lab-chip based system for automatic, continuous, on-site sampling, and sensing of heavy metals. (b) Photograph of the analyzer. (c) Polymer lab-chip with array of three sensors: (left) schematic view and (right) the photograph.¹⁴⁷

detection of heavy metals because the CNTs feature unidirectional electron transport, high electrical conductivity, and excellent strength.^{127,131} However, The CNT array suffers from the high background current and absence of functional groups for heavy metal chelation. To overcome the drawbacks, Lin et al. have sealed the side-walls of CNTs in epoxy to mitigate the current leakage and to reduce the electrode capacitance, leading to a low background current from aligned CNT forest.¹⁴⁴ This voltammetric sensor with the CNT-NEA coated with a bismuth film showed a high signal-to-noise ratio and achieved a LOD of 0.04 μ g/L for Cd²⁺.

Microfluidic Electrochemical Devices. Electrochemical sensors can be incorporated into the microfluidic channel to form a LOC device, ^{145,146} in which three electrodes (reference, counter, and working electrodes) can be integrated into a single microfluidic chamber so that the total assay process is performed by electrochemical techniques. Microfluidic electrochemical devices offer significant advantages, including (i) miniaturization of devices, (ii) enabling detection with a small sample/reagent volume, (iii) on-chip preparing sample and preconcentrating the analytes, (iv) allowing multiplexed detection in a single chip, and (v) shortening the assay time. The microfluidic electrochemical devices can be used as portable devices for field-deployable applications,147-152 Zou et al. have developed a portable analytical system using an array of disposable polymer LOCs and a continuous flow sensing method (Figure 6).¹⁴⁷ This analytical system enabled the longterm, automatic, continuous water sampling, and on-site measurement of heavy metals. More importantly, this system has a capability of monitoring multiple heavy metals using the ASV method. This system is capable of on-site measuring the Cd concentration change in the soil pore and groundwater samples.

FET SENSORS

FET sensors have been used for detection of heavy metals by utilizing the interaction between the analyte and semiconductor resistor.¹⁵³ FET sensors are capable of real-time label-free detection of heavy metals. One-dimensional and two-dimensional semiconducting nanomaterials are especially attractive to the FET sensors because they have very high surface-to-volume ratio, leading to high sensitivity.^{154,155} A Si nanowire was used to construct a FET sensor for detection of heavy metals.¹⁵⁴ This FET sensor exhibited high sensitivity with a LOD of 10^{-7} M for Hg^{2+} and 10^{-4} M for Cd^{2+} . Another example was a selective and sensitive FET sensor constructed with singlewalled carbon nanotubes (SWCNTs), which was based on the conductance change due to the selective redox reaction between SWCNTs and $Hg^{2+\,156}$ Reduction from Hg^{2+} to Hg^0 by the SWCNTs is thermodynamically favorable, while reduction of other metal ions with SWCNTs is unfavorable due to their negative potentials. This sensor exhibited a LOD of 10 nM, with a wide detection range from 10 nM to 1 mM, and excellent selectivity toward Hg²⁺ over other metal ions in both aqueous solution and drinking water.

Nanomaterials usually require the surface functionalization for specific detection of metal ions.¹⁵³ Specific detection of mercury with a FET sensor has been demonstrated by selfassembly of a 1-octadecanethiol monolayer onto graphene due to high affinity of thiol to mercury.¹⁵⁷ However, the LOD was high (~0.5 μ M). In order to improve the sensitivity, Chen et al. have used the thioglycolic acid (TGA)-functionalized reduced graphene oxide (rGO) to construct a FET sensor.¹⁵⁸ This FET sensor achieved a LOD of 2.5 × 10⁻⁸ M and responded to the analyte within a few seconds with excellent selectivity over other metal ions. The improved sensing performance was attributed to the chelation interaction of heavy metal ions with the carboxyl group of TGA, rather than the selectively binding of thiol group to mercury. In addition, the protein-functionalized rGO was employed in a FET sensor to detect various metal ions with high sensitivity.¹⁵⁹

CONCLUSIONS AND PERSPECTIVES

Enormous efforts have been made to develop heavy metal sensors in several aspects: (i) explore different signal transduction mechanisms, resulting in various optical, electrochemical, and FET sensors; (ii) utilize or develop different molecular recognition probes, including small organic molecules, DNA, aptamer, proteins, bacteria, and metals; and (iii) incorporate nanomaterials and nanostructures into sensors, leading to significant improvement in the sensing performance, especially LOD and sensitivity. However, most of the work has only demonstrated a proof-of-concept for sensors that could detect heavy metals in buffer solutions or artificial matrices. It remains a significant challenge to apply the sensors to complex real-world samples for real-time on-site detection of heavy metals. These challenges include but are not limited to the following. (i) The chemical and biological species in the realworld samples (such as river water, blood, and urine) have a severe interference on the sensing signals. Under some circumstances, the transducer of sensors suffer from fouling of organics or inorganics, leading to malfunction of the sensor. (ii) Most of the sensors reported previously were only able to detect free ions of heavy metals. Unfortunately, many heavy metals in the real-world samples are present in the form of metal-organic complexes instead of free ions. Consequently, the samples must be pretreated in a central laboratory prior to sensing. The advantages of sensors over conventional analytical techniques are their portability and on-site detection ability. If sample pretreatment has to be performed in a central laboratory prior to sensing, the enthusiasm with sensors will be severely diminished. An effective solution to these problems is to develop LOC devices that integrate sensors with microfluidics on a single chip. The microfluidic module in the LOC chip can be used to digest metal-organic complexes to release free metallic ions and separate the metallic ions from the sample matrix prior to sensing.

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Notes

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REFERENCES

(1) Duffus, J. H. Heavy metals—A meaningless term? Pure Appl. Chem. 2002, 74, 793–807.

(2) Aragay, G.; Pons, J.; Merkoçi, A. Recent trends in macro-, micro-, and nanomaterial-based tools and strategies for heavy-metal detection. *Chem. Rev.* **2011**, *111*, 3433–3458.

(3) Bings, N. H.; Bogaerts, A.; Broekaert, J. A. C. Atomic spectroscopy. *Anal. Chem.* **2006**, *78*, 3917–3946.

(4) Lorber, K. E. Monitoring of heavy metals by energy dispersive X-ray fluorescence spectrometry. *Waste Manage. Res.* **1986**, *4*, 3–13.

(5) Kunkel, R.; Manahan, S. E. Atomic absorption analysis of strong heavy metal chelating agents in water and waste water. *Anal. Chem.* **1973**, *45*, 1465–1468.

(6) Lopez-Artiguez, M.; Cameán, A.; Repetto, M. Preconcentration of heavy metals in urine and quantification by inductively coupled plasma atomic emission spectrometry. *J. Anal. Toxicol.* **1993**, *17*, 18–22.

(7) Jamali, M. R.; Assadi, Y.; Shemirani, F.; Hosseini, M. R. M.; Kozani, R. R.; Masteri-Farahani, M.; Salavati-Niasari, M. Synthesis of salicylaldehyde-modified mesoporous silica and its application as a new sorbent for separation, preconcentration and determination of uranium by inductively coupled plasma atomic emission spectrometry. *Anal. Chimi. Acta* **2006**, *579*, 68–73.

(8) Aragay, G.; Pino, F.; Merkoçi, A. Nanomaterials for sensing and destroying pesticides. *Chem. Rev.* **2012**, *112*, 5317–5338.

(9) Huang, X. J.; Choi, Y. K. Chemical sensors based on nanostructured materials. *Sens. Actuators, B* 2007, 122, 659–671.

(10) Sauer, M. Single-molecule-sensitive fluorescent sensors based on photoinduced intramolecular charge transfer. *Angew. Chem., Int. Ed.* **2003**, *42*, 1790–1793.

(11) Zhou, D.; Piper, J. D.; Abell, C.; Klenerman, D.; Kang, D. J.; Ying, L. Fluorescence resonance energy transfer between a quantum dot donor and a dye acceptor attached to DNA. *Chem. Commun.* **2005**, 38, 4807–4809.

(12) Lu, Y.; Liu, J.; Li, J.; Bruesehoff, P. J.; Pavot, C. M. B.; Brown, A. K. New highly sensitive and selective catalytic dna biosensors for metal ions. *Biosens. Bioelectron.* **2003**, *18*, 529–540.

(13) Liu, J.; Lu, Y. A DNAzyme catalytic beacon sensor for paramagnetic Cu^{2+} ions in aqueous solution with high sensitivity and selectivity. *J. Am. Chem. Soc.* **2007**, *129*, 9838–9839.

(14) Clapp, A. R.; Medintz, I. L.; Mauro, J. M.; Fisher, B. R.; Bawendi, M. G.; Mattoussi, H. Fluorescence resonance energy transfer between quantum dot donors and dye-labeled protein acceptors. *J. Am. Chem. Soc.* **2004**, *126*, 301–310.

(15) Miyake, Y.; Togashi, H.; Tashiro, M.; Yamaguchi, H.; Oda, S.; Kudo, M.; Tanaka, Y.; Kondo, Y.; Sawa, R.; Fujimoto, T.; Machinami, T.; Ono, A. Mercury^{II}-mediated formation of thymine-Hg^{II}-thymine base pairs in DNA duplexes. *J. Am. Chem. Soc.* **2006**, *128*, 2172–2173.

(16) Ono, A.; Togashi, H. Highly selective oligonucleotide-based sensor for mercury (II) in aqueous solutions. *Angew. Chem., Int. Ed.* **2004**, 43, 4300–4302.

(17) Medintz, I. L.; Clapp, A. R.; Brunel, F. M.; Tiefenbrunn, T.; Uyeda, H. T.; Chang, E. L.; Deschamps, J. R.; Dawson, P. E.; Mattoussi, H. Proteolytic activity monitored by fluorescence resonance energy transfer through quantum-dot-peptide conjugates. *Nat. Mater.* **2006**, *5*, 581–589.

(18) Li, M.; Li, R.; Li, C. M.; Wu, N. Electrochemical and optical biosensors based on nanomaterials and nanostructures: A review. *Front. Biosci.* **2011**, *3*, 1308–31.

(19) Han, M.; Gao, X.; Su, J. Z.; Nie, S. Quantum-dot-tagged microbeads for multiplexed optical coding of biomolecules. *Nat. Biotechnol.* **2001**, *19*, 631–635.

(20) Clapp, A. R.; Medintz, I. L.; Mauro, J. M.; Fisher, B. R.; Bawendi, M. G.; Mattoussi, H. Fluorescence resonance energy transfer between quantum dot donors and dye-labeled protein acceptors. *J. Am. Chem. Soc.* **2004**, *126*, 301–310.

(21) Wu, X.; Liu, H.; Liu, J.; Haley, K. N.; Treadway, J. A.; Larson, J. P.; Ge, N.; Peale, F.; Bruchez, M. P. Immunofluorescent labeling of cancer marker Her2 and other cellular targets with semiconductor quantum dots. *Nat. Biotechnol.* **2002**, *21*, 41–46.

(22) Tan, G. L.; Hömmerich, U.; Temple, D.; Wu, N. Q.; Zheng, J. G.; Loutts, G. Synthesis and optical characterization of CdTe nanocrystals prepared by ball milling process. *Scrip. Mater.* **2003**, *48*, 1469–1474.

(23) Wang, D.; Zhao, H.; Wu, N. Q.; El Khakani, A.; Ma, D. Tuning the charge transfer property of PbS-quantum dot/TiO₂-nanobelt nanohybrids via quantum confinement. *J. Phys. Chem. Lett.* **2010**, *1*, 1030–1035.

(24) Sun, Y. P.; Zhou, B.; Lin, Y.; Wang, W.; Fernando, K. S.; Pathak, P.; Meziani, M. J.; Harruff, B. A.; Wang, X.; Wang, H.; Luo, P. G.; Yang, H.; Kose, M. E.; Chen, B.; Veca, L. M.; Xie, S. Y. Quantum-sized carbon dots for bright and colorful photoluminescence. *J. Am. Chem. Soc.* **2006**, *128*, 7756–7757.

(25) Loh, K. P.; Bao, Q.; Eda, G.; Chhowalla, M. Graphene oxide as a chemically tunable platform for optical applications. *Nat. Chem.* **2010**, *2*, 1015–1024.

(26) Dong, H.; Gao, W.; Yan, F.; Ji, H.; Ju, H. Fluorescence resonance energy transfer between quantum dots and graphene oxide for sensing biomolecules. *Anal. Chem.* **2010**, *82*, 5511–5517.

(27) Wang, Y.; Bao, L.; Liu, Z.; Pang, D. W. Aptamer biosensor based on fluorescence resonance energy transfer from upconverting phosphors to carbon nanoparticles for thrombin detection in human plasma. *Anal. Chem.* **2011**, *83*, 8130–8137.

(28) Wang, L.; Yan, R.; Huo, Z.; Wang, L.; Zeng, J.; Bao, J.; Wang, X.; Peng, Q.; Li, Y. Fluorescence resonant energy transfer biosensor based on upconversion-luminescent nanoparticles. *Angew. Chem., Int. Ed.* **2005**, *44*, 6054–6057.

(29) Liu, Q.; Peng, J.; Sun, L.; Li, F. High-efficiency upconversion luminescent sensing and bioimaging of Hg(II) by chromophoric ruthenium complex-assembled nanophosphors. *ACS Nano* **2011**, *5*, 8040–8048.

(30) Li, M.; Cushing, S. K.; Wang, Q.; Shi, X.; Hornak, L. A.; Hong, Z.; Wu, N. Size-dependent energy transfer between CdSe/ZnS quantum dots and gold nanoparticles. *J. Phys. Chem. Lett.* **2011**, *2*, 2125–2129.

(31) Persson, B. N. J.; Lang, N. D. Electron-hole-pair quenching of excited states near a metal. *Phys. Rev. B* 1982, 26, 5409-5415.

(32) Yun, C. S.; Javier, A.; Jennings, T.; Fisher, M.; Hira, S.; Peterson, S.; Hopkins, B.; Reich, N. O.; Strouse, G. F. Nanometal surface energy transfer in optical rulers, breaking The FRET barrier. *J. Am. Chem. Soc.* **2005**, *127*, 3115–3119.

(33) Jennings, T. L.; Singh, M. P.; Strouse, G. F. Fluorescent lifetime quenching near d = 1.5 nm gold nanoparticles: Probing NSET validity. *J. Am. Chem. Soc.* **2006**, *128*, 5462–5467.

(34) Ray, P. C.; Fortner, A.; Darbha, G. K. Gold nanoparticle based FRET assay for the detection of DNA cleavage. *J. Phys. Chem. B* 2006, 110, 20745–20748.

(35) Mayilo, S.; Kloster, M. A.; Wunderlich, M.; Lutich, A.; Klar, T. A.; Nichtl, A.; Kürzinger, K.; Stefani, F. D.; Feldmann, J. Long-range fluorescence quenching by gold nanoparticles in a sandwich immunoassay for cardiac troponin T. *Nano Lett.* **2009**, *9*, 4558–4563. (36) De, M.; Ghosh, P. S.; Rotello, V. M. Applications of

nanoparticles in biology. Adv. Mater. 2008, 20, 4225–4241.

(37) Pons, T.; Medintz, I. L.; Sapsford, K. E.; Higashiya, S.; Grimes, A. F.; Doug, S.; Mattoussi, H. On the quenching of semiconductor quantum dot photoluminescence by proximal gold nanoparticles. *Nano Lett.* **2007**, *7*, 3157–3164.

(38) Li, M.; Wang, Q.; Shi, X.; Hornak, L. A.; Wu, N. Detection of mercury (II) by quantum dot/DNA/gold nanoparticle ensemble based nanosensor via nanometal surface energy transfer. *Anal. Chem.* **2011**, 83, 7061–7065.

(39) Darbha, G. K.; Ray, A.; Ray, P. C. Gold nanoparticle-based miniaturized nanomaterial surface energy transfer probe for rapid and ultrasensitive detection of mercury in soil, water, and fish. *ACS Nano* **2007**, *1*, 208–214.

(40) Li, M.; Zhou, X.; Guo, S.; Wu, N. Detection of lead (II) with a "turn-on" fluorescent biosensor based on energy transfer from CdSe/ ZnS quantum dots to graphene oxide. *Biosens. Bioelectron.* **2013**, 43, 69–74.

(41) Swathi, R. S.; Sebastian, K. L. Resonance energy transfer from a dye molecule to graphene. J. Chem. Phys. 2008, 129, 054703.

(42) Swathi, R. S.; Sebastian, K. L. Long range resonance energy transfer from a dye molecule to graphene has $(distance)^{-4}$ dependence. *J. Chem. Phys.* **2009**, *130*, 086101.

(43) Li, M.; Cushing, S. K.; Zhou, X.; Guo, S.; Wu, N. Q. Fingerprinting photoluminescence of functional groups in graphene oxide. *J. Mater. Chem.* **2012**, *22*, 23374–23379.

(44) Jung, J. H.; Cheon, D. S.; Liu, F.; Lee, K. B.; Seo, T. S. A graphene oxide based immuno-biosensor for pathogen detection. *Angew. Chem., Int. Ed.* **2010**, *49*, 5708–5711.

(45) Chen, D.; Feng, H.; Li, J. Grapheneoxide: Preparation, functionalization, and electrochemical applications. *Chem. Rev.* 2012, *112*, 6027–6053.

(46) Lee, J. S.; Joung, H. A.; Kim, M. G.; Park, C. B. Graphene-based chemiluminescence resonance energy transfer for homogeneous immunoassay. *ACS Nano* **2012**, *6*, 2978–2983.

(47) Zhang, M.; Yin, B. C.; Tan, W.; Ye, B. C. A versatile graphenebased fluorescence "on/off" switch for multiplex detection of various targets. *Biosens. Bioelectron.* **2011**, *26*, 3260–3265.

(48) Chen, K.; Lu, G.; Chang, J.; Mao, S.; Yu, K.; Cui, S.; Chen, J. Hg (II) ion detection using thermally reduced graphene oxide decorated with functionalized gold nanoparticles. *Anal. Chem.* **2012**, *84*, 4057–4062.

(49) Chang, H.; Tang, L.; Wang, Y.; Jiang, J.; Li, J. Graphene fluorescence resonance energy transfer aptasensor for the thrombin detection. *Anal. Chem.* **2010**, *82*, 2341–2346.

(50) Li, J. L.; Bao, H. C.; Hou, X. L.; Sun, L.; Wang, X. G.; Gu, M. Graphene oxide nanoparticles as a nonbleaching optical probe for twophoton luminescence imaging and cell therapy. *Angew. Chem., Int. Ed.* **2012**, *51*, 1830–1834.

(51) Wen, Y.; Xing, F.; He, S.; Song, S.; Wang, L.; Long, Y.; Li, D.; Fan, C. A graphene-based fluorescent nanoprobe for silver (I) lins detection by using graphene oxide and a silver-specific oligonucleotide. *Chem. Commun.* **2010**, *46*, 2596–2598.

(52) Zhao, X. H.; Kong, R. M.; Zhang, X. B.; Meng, H. M.; Liu, W. N.; Tan, W.; Shen, G. L; Yu, R. Q. Graphene–DNAzyme based biosensor for amplified fluorescence "turn-on" detection of Pb²⁺ with a high selectivity. *Anal. Chem.* **2011**, *83*, 5062–5066.

(53) Freeman, R.; Liu, X.; Willner, I. Chemiluminescent and chemiluminescence resonance energy transfer (CRET) detection of DNA, metal ions, and aptamer–substrate complexes using hemin/G-quadruplexes and CdSe/ZnS quantum dots. J. Am. Chem. Soc. 2011, 133, 11597–11604.

(54) Freeman, R.; Willner, B.; Willner, I. Integrated biomolecule– quantum dot hybrid systems for bioanalytical applications. *J. Phys. Chem. Lett.* **2011**, 2, 2667–2677.

(55) Freeman, R.; Finder, T.; Willner, I. Multiplexed analysis of Hg^{2+} and Ag^+ ions by nucleic acid functionalized CdSe/ZnS quantum dots and their use for logic gate operations. *Angew. Chem., Int. Ed.* **2009**, *48*, 7818–7821.

(56) Li, M.; Zhou, X.; Ding, W.; Guo, S.; Wu, N. Fluorescent aptamer-functionalized graphene oxide biosensor for label-free detection of mercury (II). *Biosens. Bioelectron.* **2013**, *41*, 889–893.

(57) Gao, Y. Q.; Georgievskii, Y.; Marcus, R. A. On the theory of electron transfer reactions at semiconductor electrode/liquid interfaces. J. Chem. Phys. 2000, 112, 3358–3369.

(58) Dexter, D. L. A theory of sensitized luminescence in solids. J. Chem. Phys. 1953, 21, 836–850.

(59) Lakowicz, J. R. Principles of Fluorescence Spectroscopy, 3rd ed.; Spinger Academic: New York, 2006.

(60) Li, M.; Cushing, S. K.; Zhang, J.; Lankford, J.; Aguilar, Z. P.; Ma, D.; Wu, N. Shape-dependent surface-enhanced raman scattering in gold–raman-probe–silica sandwiched nanoparticles for biocompatible applications. *Nanotechnology* **2012**, *23*, 115501.

(61) Mayer, K. M.; Hafner, J. H. Localized surface plasmon resonance sensors. *Chem. Rev.* 2011, 111, 3828–3857.

(62) Xu, X.; Daniel, W. L.; Wei, W.; Mirkin, C. A. Colorimetric Cu²⁺ detection using DNA-modified gold-nanoparticle aggregates as probes and click chemistry. *Small* **2010**, *6*, 623–626.

(63) Zhang, X. B.; Kong, R. M.; Lu, Y. Metal ion sensors based on DNAzymes and related DNA molecules. *Ann. Rev. Anal. Chem.* 2011, *4*, 105.

(64) Liu, J.; Lu, Y. Accelerated color change of gold nanoparticles assembled by DNAzymes for simple and fast colorimetric Pb²⁺ detection. *J. Am. Chem. Soc.* **2004**, *126*, 12298–12305.

(65) Liu, J.; Lu, Y. A Colorimetric Lead Biosensor Using DNAzyme-Directed Assembly of Gold Nanoparticles. J. Am. Chem. Soc. 2003, 125, 6642–6643.

(66) Slocik, J. M.; Zabinski, J. S.; Phillips, D. M.; Naik, R. R. Colorimetric response of peptide-functionalized gold nanoparticles to metal ions. *Small* **2008**, *4*, 548–551.

(67) Darbha, G. K.; Singh, A. K.; Rai, U. S.; Yu, E.; Yu, H.; Chandra Ray, P. Selective detection of mercury (II) ion using nonlinear optical properties of gold nanoparticles. *J. Am. Chem. Soc.* **2008**, *130*, 8038–8043.

(68) Kim, Y.; Johnson, R. C.; Hupp, J. T. Gold nanoparticle-based sensing of "spectroscopically silent" heavy metal ions. *Nano Lett.* **2001**, *1*, 165–167.

(69) Storhoff, J. J.; Elghanian, R.; Mucic, R. C.; Mirkin, C. A.; Letsinger, R. L. One-pot colorimetric differentiation of polynucleotides with single base imperfections using gold nanoparticle probes. *J. Am. Chem. Soc.* **1998**, *120*, 1959–1964.

(70) Lee, J. S.; Han, M. S.; Mirkin, C. A. Colorimetric detection of mercuric ion (Hg^{2+}) in aqueous media using DNA-functionalized gold nanoparticles. *Angew. Chem.* **2007**, *119*, 4171–4174.

(71) Li, T.; Wang, E.; Dong, S. Lead (II)-induced allosteric Gquadruplex DNAzyme as a colorimetric and chemiluminescence sensor for highly sensitive and selective Pb²⁺ detection. *Anal. Chem.* **2010**, *82*, 1515–1520.

(72) Liu, J.; Lu, Y. Stimuli-responsive disassembly of nanoparticle aggregates for light-up colorimetric sensing. J. Am. Chem. Soc. 2005, 127, 12677–12683.

(73) Chai, F.; Wang, C.; Wang, T.; Li, L.; Su, Z. Colorimetric detection of Pb^{2+} using glutathione functionalized gold nanoparticles. *ACS Appl. Mater. Interfaces* **2010**, *2*, 1466–1470.

(74) He, X.; Liu, H.; Li, Y.; Wang, S.; Wang, N.; Xiao, J.; Xu, X.; Zhu, D. Gold nanoparticle-based fluorometric and colorimetric sensing of copper (II) ions. *Adv. Mater.* **2005**, *17*, 2811–2815.

(75) Kalluri, J. R.; Arbneshi, T.; Afrin Khan, S.; Neely, A.; Candice, P.; Varisli, B.; Washinton, M.; McAfee, S.; Robinson, B.; Banerjee, S.; Singh, A. K.; Senapati, D.; Ray, P. C. Use of gold nanoparticles in a simple colorimetric and ultrasensitive dynamic light scattering assay: Selective detection of arsenic in groundwater. *Angew. Chem., Int. Ed.* **2009**, *48*, 9668–9671.

(76) Xue, X.; Wang, F.; Liu, X. One-step, room temperature, colorimetric detection of mercury (Hg^{2+}) using DNA/nanoparticle conjugates. J. Am. Chem. Soc. **2008**, 130, 3244–3245.

(77) Yu, C. J.; Tseng, W. L. Colorimetric detection of mercury (II) in a high-salinity solution using gold nanoparticles capped with 3mercaptopropionate acid and adenosine monophosphate. *Langmuir* **2008**, *24*, 12717–12722.

(78) Pitarke, J. M.; Silkin, V. M.; Chulkov, E. V.; Echenique, P. M. Theory of surface plasmons and surface-plasmon polaritons. *Rep. Prog. Phys.* **2006**, *70*, 1.

(79) Homola, J.; Yee, S. S.; Gauglitz, G. Surface plasmon resonance sensors: Review. *Sens. Actuators, B* **1999**, *54*, 3–15.

(80) Gao, S.; Koshizaki, N.; Koyama, E.; Tokuhisa, H.; Sasaki, T.; Kim, J. K.; Cho, Y.; Kim, D. S.; Shimizu, Y. Innovative platform for transmission localized surface plasmon transducers and its application in detecting heavy metal Pd (II). *Anal. Chem.* **2009**, *81*, 7703–7712.

(81) Forzani, E. S.; Zhang, H.; Chen, W.; Tao, N. Detection of heavy metal ions in drinking water using a high-resolution differential surface plasmon resonance sensor. *Environ. Sci. Technol.* **2005**, *39*, 1257–1262.

(82) Li, M.; Cushing, S. K.; Liang, H.; Suri, S.; Ma, D.; Wu, N. Q. Plasmonic nanorice antenna on triangle nano-array for surfaceenhanced raman scattering detection of hepatitis B virus DNA. *Anal. Chem.* **2013**, *85*, 2072–2078.

(83) Li, M.; Zhang, J.; Suri, S.; Sooter, L. J.; Ma, D.; Wu, N. Q. Detection of adenosine triphosphate with an aptamer bosensor based on surface-enhanced raman scattering. *Anal. Chem.* **2012**, *84*, 2837–2842.

(84) Jimenez de Aberasturi, D.; Montenegro, J. M.; Ruiz de Larramendi, I.; Rojo, T.; Klar, T. A.; Alvarez-Puebla, R.; Liz-Marzán, (85) Li, J.; Chen, L.; Lou, T.; Wang, Y. Highly sensitive SERS detection of As^{3+} ions in aqueous media using glutathione functionalized silver nanoparticles. *ACS Appl. Mater. Interfaces* **2012**, *3*, 3936–3941.

(86) Han, D.; Lim, S. Y.; Kim, B. J.; Piao, L.; Chung, T. D. Mercury(II) detection by SERS based on a single gold microshell. *Chem. Commun.* **2010**, *46*, 5587–5589.

(87) Mulvihill, M.; Tao, A.; Benjauthrit, K.; Arnold, J.; Yang, P. Surface-enhanced Raman spectroscopy for trace arsenic detection in contaminated water. *Angew. Chem., Int. Ed.* **2008**, 47, 6456–6460.

(88) Feilchenfeldt, H.; Siiman, O. Surface Raman excitation and enhancement profiles for chromate, molybdate, and tungstate on colloidal silver. *J. Phys. Chem.* **1986**, *90*, 2168–2173.

(89) Bao, L.; Mahurin, S. M.; Haire, R. G.; Dai, S. Silver-doped solgel film as a surface-enhanced raman scattering substrate for detection of uranyl and neptunyl Ions. *Anal. Chem.* **2003**, *75*, 6614–6620.

(90) Gu, B.; Ruan, C. Determination of technetium and its speciation by surface-enhanced Raman spectroscopy. *Anal. Chem.* **2007**, *79*, 2341–2345.

(91) Ravindranath, S. P.; Henne, K. L.; Thompson, D. K.; Irudayaraj, J. Raman chemical imaging of chromate reduction sites in a single bacterium using intracellularly grown gold nanoislands. *ACS Nano* **2011**, *5*, 4729–4736.

(92) Fan, X.; White, I. M. Optofluidic microsystems for chemical and biological analysis. *Nat. Photonics* **2011**, *5*, 591–597.

(93) Faye, D.; Zhang, H.; Lefevre, J. P.; Bell, J.; Delaire, J.; Leray, I. Mercury detection in a microfluidic device by using a molecular sensor soluble in organoaqueous solvent. *Photochem. Photobiol. Sci.* **2012**, *11*, 1737–1743.

(94) Chang, I. H.; Tulock, J. J.; Liu, J.; Kim, W. S.; Cannon, D. M., Jr.; Lu, Y.; Bohn, P. W.; Sweedler, J. V.; Cropek, D. M. Miniaturized lead sensor based on lead-specific DNAzyme in a nanocapillary interconnected microfluidic device. *Environ. Sci. Technol.* **2005**, *39*, 3756–3761.

(95) Lafleur, J.; Senkbeil, S.; Jensen, T. G.; Kutter, J. P. Gold nanoparticle-based optical microfluidic sensors for analysis of environmental pollutants. *Lab Chip* **2012**, *12*, 4651–4656.

(96) Kou, S.; Nam, S. W.; Shumi, W.; Lee, M. H.; Bae, S. W.; Du, J.; Kim, J. S.; Hong, J. I.; Peng, X.; Yoon, J.; Park, S. Microfluidic detection of multiple heavy metal ions using fluorescent chemosensors. *Bull. Korean Chem. Soc.* **2009**, *30*, 1173–1176.

(97) Wang, G.; Lim, C.; Chen, L.; Chon, H.; Choo, J.; Hong, J.; demello, A. J. Surface-enhanced Raman scattering in nanoliter droplets: Towards high-sensitivity detection of mercury (II) ions. *Anal. Bioanal. Chem.* **2009**, *394*, 1827–1832.

(98) Lee, S. J.; Moskovits, M. Visualizing chromatographic separation of metal ions on a surface-enhanced Raman active medium. *Nano Lett.* **2011**, *11*, 145–150.

(99) Yang, X.; Shi, C.; Wheeler, D.; Newhouse, R.; Chen, B.; Zhang, J. Z.; Gu, C. High-sensitivity molecular sensing using hollow-core photonic crystal fiber and surface-enhanced Raman scattering. *J. Opt. Soc. Am. A* **2010**, *27*, 977–984.

(100) Khaing Oo, M. K.; Han, Y.; Kanka, J.; Sukhishvili, S.; Du, H. Structure fits the purpose: Photonic crystal fibers for evanescent-field surface-enhanced Ramanspectroscopy. *Opt. Lett.* **2010**, *35*, 466–469.

(101) Wang, M.; Jing, N.; Chou, I. H.; Cote, G. L.; Kameoka, J. An Optofluidic Device for Surface Enhanced Raman Spectroscopy. *Lab Chip* **2007**, *7*, 630–632.

(102) Liu, J.; White, I.; DeVoe, D. L. Nanoparticle-functionalized porous polymer monolith detection elements for surface-enhanced Raman scattering. *Anal. Chem.* **2011**, *83*, 2119–2124.

(103) Kimmel, D. W.; LeBlanc, G.; Meschievitz, M. E.; Cliffel, D. E. Electrochemical sensors and biosensors. *Anal. Chem.* **2012**, *84*, 685–707.

(104) Krawczyński vel Krawczyk, T.; Moszczyńska, M.; Trojanowicz, M. Inhibitive determination of mercury and other metal ions by potentiometric urea biosensor. *Biosens. Bioelectron.* **2000**, *15*, 681–691.

(105) Li, X. G.; Feng, H.; Huang, M. R.; Gu, G. L.; Moloney, M. G. Ultrasensitive Pb (II) potentiometric sensor based on copolyaniline nanoparticles in a plasticizer-free membrane with a long lifetime. *Anal. Chem.* **2012**, *84*, 134–140.

(106) Shtoyko, T.; Maghasi, A. T.; Richardson, J. N.; Seliskar, C. J.; Heineman, W. R. Spectroelectrochemical sensing based on attenuated total internal reflectance stripping voltammetry. 1. Determination of lead and cadmium. *Anal. Chem.* **2003**, *75*, 4585–4590.

(107) Rassaei, L.; Marken, F.; Sillanpää, M.; Amiri, M.; Cirtiu, C. M.; Sillanpää, M. Nanoparticles in electrochemical sensors for environmental monitoring. *Trends Anal. Chem.* **2011**, *30*, 1704–1714.

(108) Kuila, T.; Bose, S.; Khanra, P.; Mishra, A. K.; Kim, N. H.; Lee, J. H. Recent advances in graphene-based biosensors. *Biosens. Bioelectron.* **2011**, *26*, 4637–4648.

(109) Tarley, C. R. T.; Santos, V. S.; Baêta, B. E. L.; Pereira, A. C.; Kubota, L. T. Simultaneous determination of zinc, cadmium and lead in environmental water samples by potentiometric stripping analysis (PSA) using multiwalled carbon nanotube electrode. *J. Hazard. Mater.* **2009**, *169*, 256–262.

(110) Aragay, G.; Puig-Font, A.; Cadevall, M.; Merkoçi, A. Surface characterizations of mercury-based electrodes with the resulting micro and nano amalgam wires and spheres formations may reveal both gained sensitivity and faced nonstability in heavy metal detection. *J. Phys. Chem. C* 2010, *114*, 9049–9055.

(111) Wang, J.; Lu, J.; Hocevar, S. B.; Farias, P. A.; Ogorevc, B. Bismuth-coated carbon electrodes for anodic stripping voltammetry. *Anal. Chem.* **2000**, *72*, 3218–3222.

(112) Švancara, I.; Prior, C.; Hočevar, S. B.; Wang, J. A decade with bismuth-based electrodes in electroanalysis. *Electroanalysis* **2010**, *22*, 1405–1420.

(113) Pan, D.; Wang, Y.; Chen, Z.; Lou, T.; Qin, W. Nanomaterial/ ionophore-based electrode for anodic stripping voltammetric determination of lead: An electrochemical sensing platform toward heavy metals. *Anal. Chem.* **2009**, *81*, 5088–5094.

(114) Manivannan, A.; Seehra, M. S.; Tryk, D. A.; Fujishima, A. Electrochemical detection of ionic mercury at boron-doped diamond electrodes. *Anal. Lett.* **2002**, *35*, 355–368.

(115) Gao, X.; Wei, W.; Yang, L.; Yin, T.; Wang, Y. Simultaneous determination of lead, copper, and mercury free from macromolecule contaminants by square wave stripping voltammetry. *Anal. Lett.* **2005**, 38, 2327–2343.

(116) Abollino, O.; Giacomino, A.; Malandrino, M.; Piscionieri, G.; Mentasti, E. Determination of mercury by anodic stripping voltammetry with a gold nanoparticle-modified glassy carbon electrode. *Electroanalysis* **2008**, *20*, 75–83.

(117) Dai, X.; Compton, R. G. Gold nanoparticle modified electrodes show a reduced interference by Cu (II) in the detection of As (III) using anodic stripping voltammetry. *Electroanalysis* **2005**, *17*, 1325– 1330.

(118) Khairy, M.; Kampouris, D. K.; Kadara, R. O.; Banks, C. E. Gold nanoparticle modified screen printed electrodes for the trace sensing of arsenic (III) in the presence of copper (II). *Electroanalysis* **2010**, *22*, 2496–2501.

(119) Oyama, M. Recent nanoarchitectures in metal nanoparticlemodified electrodes for electroanalysis. *Anal. Sci.* **2010**, *26*, 1–12.

(120) de la Escosura-Muñiz, A.; Ambrosi, A.; Merkoçi, A. Electrochemical analysis with nanoparticle-based biosystems. *Trends Anal.Chem.* **2008**, *27*, 568–584.

(121) Aragay, G.; Merkoci, A. Nanomaterials application in electrochemical detection of heavy metals. *Electrochim. Acta* 2012, *84*, 49–61.

(122) Khairy, M.; Choudry, N. A.; Ouasti, M.; Kampouris, D. K.; Kadara, R. O.; Banks, C. E. Gold nanoparticle ensembles allow mechanistic insights into electrochemical processes. *ChemPhysChem* **2010**, *11*, 875–879.

(123) Majid, E.; Hrapovic, S.; Liu, Y.; Keith, B.; Luong, J. H. Electrochemical determination of arsenite using a gold nanoparticle modified glassy carbon electrode and flow analysis. *Anal. Chem.* **2006**, 78, 762–769.

(124) Dai, X.; Compton, R. G. Gold nanoparticle modified electrodes show a reduced interference by Cu(II) in the detection of As(III) using anodic stripping voltammetry. *Electroanalysis* **2005**, *17*, 1325– 1330.

(125) Lee, G.-J.; Kim, C. K.; Lee, M. K.; Rhee, C. K. Simultaneous voltammetric determination of Zn, Cd and Pb at Bismuth Nanopowder Electrodes with Various Particle Size Distributions. *Electroanalysis* **2010**, *22*, 530–535.

(126) Shao, Y.; Wang, J.; Wu, H.; Liu, J.; Aksay, I. A.; Lin, Y. Graphene based electrochemical sensors and biosensors: A review. *Electroanalysis* **2010**, *22*, 1027–1036.

(127) Wanekaya, A. K. Applications of nanoscale carbon-based materials in heavy metal sensing and detection. *Analyst* 2011, 136, 4383-4391.

(128) Bui, M. P.; Li, C. A.; Han, K. N.; Pham, X. H.; Seong, G. H. Electrochemical determination of cadmium and lead on pristine singlewalled carbon nanotube electrodes. *Anal. Sci.* **2012**, *28*, 699–704.

(129) Chen, K.; Lu, G.; Chang, J.; Mao, S.; Yu, K.; Cui, S.; Chen, J. Hg (II) ion detection using thermally reduced graphene oxide decorated with functionalized gold nanoparticles. *Anal. Chem.* **2012**, *84*, 4057–4062.

(130) Wang, D. W.; Li, F.; Zhao, J.; Ren, W.; Chen, Z. G.; Tan, J.; Wu, Z. S.; Gentle, I.; Lu, G. Q.; Cheng, H. M. Fabrication of graphene/polyaniline composite paper via in situ anodic electro-polymerization for high-performance flexible electrode. *ACS Nano* **2009**, *3*, 1745–1752.

(131) Musameh, M. M.; Hickey, M.; Kyratzis, I. L. Carbon nanotubebased extraction and electrochemical detection of heavy metals. *Res. Chem. Intermed.* **2011**, *37*, 675–689.

(132) Wang, L.; Wang, X.; Shi, G.; Peng, C.; Ding, Y. Thiacalixarene covalently functionalized multiwalled carbon nanotubes as chemically modified electrode material for detection of ultratrace Pb²⁺ ions. *Anal. Chem.* **2012**, *84*, 10560–10567.

(133) Morton, J.; Havens, N.; Mugweru, A.; Wanekaya, A. K. Detection of trace heavy metal ions using carbon nanotube-modified electrodes. *Electroanalysis* **2009**, *21*, 1597–1603.

(134) Glaspell, G.; Hassan, H. M.; Elzatahry, A.; Fuoco, L.; Radwan, N. R.; El-Shall, M. S. Nanocatalysis on tailored shape supports: Au and Pd nanoparticles supported on MgO nanocubes and ZnO nanobelts. *J. Phys. Chem. B* **2006**, *110*, 21387–21393.

(135) Wei, Y.; Gao, C.; Meng, F. L.; Li, H. H.; Wang, L.; Liu, J. H.; Huang, X. J. SnO_2 /reduced graphene oxide nanocomposite for the simultaneous electrochemical detection of cadmium (II), lead (II), copper (II), and mercury (II): An interesting favorable mutual interference. J. Phys. Chem. C **2011**, 116, 1034–1041.

(136) Mališić, M.; Janošević, A.; Paunlović, Š. B.; Stojković, I.; Ćirić-Marjanović, G. Exploration of MnO₂/carbon composites and their application to simultaneous electroanalytical determination of Pb(II) and Cd(II). *Electrochim. Acta* **2012**, *74*, 158–164.

(137) Liu, X.; Hu, Q.; Fang, Z.; Zhang, X.; Zhang, B. Magnetic chitosan nanocomposites: A useful recyclable tool for heavy metal ion removal. *Langmuir* **2008**, *25*, 3–8.

(138) Li, H.; Wu, N. Q. A large-area nanoscale gold hemisphere pattern as a nanoelectrode array. *Nanotechnology* **2008**, *19*, 275301.

(139) Ordeig, O.; del Campo, J.; Munoz, F. X.; Banks, C. E.; Compton, R. G. Electroanalysis utilizing amperometric microdisk electrode arrays. *Electroanalysis* **2007**, *19*, 1973–1986.

(140) Huang, X. J.; O'Mahony, A. M.; Compton, R. G. Microelectrode arrays for electrochemistry: Approaches tofabrication. *Small* **2009**, *5*, 776–788.

(141) Ugo, P.; Moretto, L. M.; Vezzà, F. Ionomer-coated electrodes and nanoelectrode ensembles as electrochemical environmental sensors: recent advances and prospects. *ChemPhysChem* 2002, *3*, 917–925.

(142) Xie, X.; Stueben, D.; Berner, Z. The application of microelectrodes for the measurements of trace metals in water. *Anal. Lett.* **2005**, *38*, 2281–2300.

(143) Ordeig, O.; Banks, C. E.; del Campo, J.; Muñoz, F. X.; Compton, R. G. Trace detection of mercury (II) using gold ultramicroelectrode arrays. *Electroanalysis* **2006**, *18*, 573–578.

(144) Liu, G.; Lin, Y.; Tu, Y.; Ren, Z. Ultrasensitive voltammetric detection of trace heavy metal ions using carbon nanotube nanoelectrode array. *Analyst* **2005**, *130*, 1098–1101.

(145) Jang, A.; Zou, Z.; Lee, K. K.; Ahn, C. H.; Bishop, P. L. State-of-the-art lab chip sensors for environmental water monitoring. *Meas. Sci. Technol.* **2011**, *22*, 032001.

(146) Du, Y.; Chen, C.; Zhou, M.; Dong, S.; Wang, E. Microfluidic electrochemical aptameric assay integrated on-chip: A potentially convenient sensing platform for the amplified and multiplex analysis of small molecules. *Anal. Chem.* **2011**, *83*, 1523–1529.

(147) Zou, Z.; Jang, A.; MacKnight, E. T.; Wu, P. M.; Do, J.; Shim, J. S.; Bishop, P. L.; Ahn, C. H. An on-site heavy metal analyzer with polymer lab-on-a-chips for continuous sampling and monitoring. *IEEE Sensors J.* **2009**, *9*, 586–594.

(148) Zou, Z.; Jang, A.; MacKnight, E.; Wu, P. M.; Do, J.; Bishop, P. L.; Ahn, C. H. Environmentally friendly disposable sensors with microfabricated on-chip planar bismuth electrode for *in situ* heavy metal ions measurement. *Sens. Actuators, B* **2008**, *134*, 18–24.

(149) Chen, C.; Zhang, J.; Du, Y.; Yang, X.; Wang, E. Microfabricated on-chip integrated Au–Ag–Au three-electrode system for in situ mercury ion determination. *Analyst* **2010**, *135*, 1010–1014.

(150) Jung, W.; Jang, A.; Bishop, P. L.; Ahn, C. H. A polymer lab chip sensor with microfabricated planar silver electrode for continuous and on-site heavy metal measurement. *Sens. Actuators, B* **2011**, *155*, 145–153.

(151) Lin, Y.; Timchalk, C. A.; Matson, D. W.; Wu, H.; Thrall, K. D. Integrated microfluidics/electrochemical sensor system for monitoring of environmental exposures to lead and chlorophenols. *Biomed. Microdev.* **2001**, *3*, 331–338.

(152) Jang, A.; Zou, Z.; Lee, K. K.; Ahn, C. H.; Bishop, P. L. Potentiometric and voltammetric polymer lab chip sensors for determination of nitrate, pH and Cd (II) in water. *Talanta* **2010**, 83, 1–8.

(153) Chen, K. I.; Li, B. R.; Chen, Y. T. Silicon nanowire field-effect transistor-based biosensors for biomedical diagnosis and cellular recording investigation. *Nano Today* **2011**, *6*, 131–154.

(154) Luo, L.; Jie, J.; Zhang, W.; He, Z.; Wang, J.; Yuan, G.; Zhang, W.; Wu, L. C. M.; Lee, S. T. Synthesis of semiconductor micro/ nanowires and their applications in devices. *Appl. Phys. Lett.* **2009**, *94*, 193101.

(155) Sudibya, H. G.; He, Q.; Zhang, H.; Chen, P. Electrical detection of metal ions using field-effect transistors based on micropatterned reduced graphene oxide films. *ACS Nano* **2011**, *5*, 1990–1994.

(156) Kim, T. H.; Lee, J.; Hong, S. Highly selective environmental nanosensors based on anomalous response of carbon nanotube conductance to mercury ions. *J. Phys. Chem. C* **2009**, *113*, 19393–19396.

(157) Zhang, T.; Cheng, Z.; Wang, Y.; Li, Z.; Wang, C.; Li, Y.; Fang, Y. Self-assembled 1-octadecanethiol monolayers on graphene for mercury detection. *Nano Lett.* **2010**, *10*, 4738–4741.

(158) Chen, K.; Lu, G.; Chang, J.; Mao, S.; Yu, K.; Cui, S.; Chen, J. hg (ii) ion detection using thermally reduced graphene oxide decorated with functionalized gold nanoparticles. *Anal. Chem.* **2012**, *84*, 4057–4062.

(159) Sudibya, H. G.; He, Q.; Zhang, H.; Chen, P. Electrical detection of metal ions using field-effect transistors based on micropatterned reduced graphene oxide films. *ACS Nano* **2011**, *5*, 1990–1994.